Application of new technologies in environmental infection control in a regional hospital in Hong Kong



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27 February 2012 Commissioned Training on Disinfection and Sterilization

Increasing body of evidence showing the importance of environment in transmission of HCA pathogens from infected patients to susceptible patients:

- Contamination of hosp env
- Survival of pathogens on surface
- Indirect transmission: acquisition on HCW hands after contact w/ contaminated env surfaces
- Direct transmission: patient acquire pathogens after contact w/ env
- Env cleaning reduces transmission of pathogens

Persistence of clinically relevant bacteria on dry inanimate surfaces

- Acinetobacter spp.: 3 days 5 months
- Clostridium difficile (spores): 5 months
- Enterococcus spp. including VRE and VSE: 5 days 4 months
- Staphylococcus aureus, including MRSA: 7 days 7 months
- Norovirus and feline calicivirus: 8 hours 7 days

• WHO Infection Control Toolkit (HA IDC)

Acquisition of MRSA on hands after contact w/ skin (40%) and env (45%)

- Stiefel U et al. SHEA 2010
- Donskey CJ et al. NEJM 2009

No difference in <u>skin & env contamination</u>, and <u>hand acquisition</u> of MRSA after skin contact w/ MRSA carriers identified *clinically* Vs thru' *active surveillance Chang S. et al. CID 2009*



Figure 10.2. Picture of room noting some high touch surfaces and items.

5 Moments of Hand Hygiene (WHO)



Even with high compliance to HH, crosstransmission still occurs

 Pittet D et al. Ann Intern Med 1999;130: 126-30, 159:821-6 Problems in manual cleansing in clinical wards: How clean are they?

Assign Responsibility for Cleaning Procedures

- Housekeepers and nursing staff often do not agree on who should clean what
- Housekeepers do not always understand
 - Which detergent/disinfectant to use
 - What concentration should be used
 - What contact times are recommended
 - How often to change cleaning cloths/mop heads
- Develop policies regarding who should clean what

Dumigan DG et al. AJIC 2010 (in press)



Ways out

- To improve environmental surface manual cleaning
 - What items have been missed
 - How clean are they if not missed
- > To look for better surface disinfectants

Ideal tools for checking surface cleanliness

- Sensitive
- Specific
- Quantitative Vs qualitative
- > Objective
- > Available interpretative standard
- > Reproducible
- No Hawthorne effect
- User friendly
- > Quick: real-time
- Cheap
- Safe

Checking surface cleanliness

- Indications: routine Vs ad hoc
 - Training
 - Quality assurance
 - Outbreak investigations

How to monitor the effectiveness of surface disinfection?

Methods for assessing cleaning practices

- Visual inspection of surfaces
- Observation of housekeeper techniques
- Aerobic colony count (ACC)
- Fluorescent marker system
- Adeosine triphosphate (ATP) bioluminescence

Improving cleaning & disinfection: How to monitor the effectiveness of surface disinfection?

> ACC

- Moistened swab inoculated onto agar +/broth enrichment
- RODAC (replicate organism detection and counting) plates
- ACC using cellulose sponges
- No standard methods
- No accepted criteria



Fluorescent marker system

DAZO® Fluorescent Marking Gel (Ecolab)





Trials at PMH



清潔成績圖示



Site of spot	Uncleaned surfaces still	Site of spot	Cleaned surfa	ed surfaces showed		
check	stained by fluorescence	check	no fluorescene	fluorescence		
Bed end		Bedside rail				
Bedside	S D	Remote	added of a	and the second day in the line		
trolley	J	control (Bed	and the second se	-		
drawer		2)				
Bedside			nitional control			
noney				清潔能力評估表		
surface			日期:	時間:		
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			床頭位置	Maria		
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Appendix 9(B) 清漆能力	評估表
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部門:	病房:
清潔人員:	評估員:
age装饰故意人位田: 每 / 方	

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Collaboration with HKUST

Secondary school project on fluorescent dye pen for hygiene monitoring



ATP bioluminescence

 surrogate marker of bacterial load
 quick, easy, sensitive
 recently proposed as one of the standards for surface hygiene in hospitals in UK

How clean in targeted wards – using 3M ATP Method



Date : 25 June 2009 Venue: G615 W C TANG WM ICT&IDC Infection Control Team

- Use 3M Clean-Trace ATP Surface Hygiene Test
- Swab all sampling areas, common touching area in ward setting
- o Review the sampling result

Sampling







Floor Plan



Sampling locations



BP cuff



Bed side Basin 1



bedside rail









locker

Dressing trolley





Dressing tray

Bottom shelf of dressing tray

Patient Toilet and shower head



Basin handle

Shower tab

Bathroom handle

Toilet basins

o Patient toilet basin handle



Control ward



Control ward

o Patient toilet basin handle



Data review between two wards

<u>5m cient-fracesultate A17 Results - FRA</u>										
br <i>é</i> W	Ward	Ward	Ward	Ward	(Control)	(Control)	(Control)			
	23/6/09 at 10am	23/6/09 st 4pm	24/6/09 at 11am	25/6/09 at 11am	23/6/09 st 4pm	24/6/09 at 10am	24/6/09 at 10am			
(wobbed sites	(FLU)	(FLU)	After cleansing before patient use (RLU)	Cleansing under ES spervisor's supervision (RLU)	(TLT)	(FLU)	After cleansing before patient use (RLU)			
Basin1	226	184		19	63	224				
Basin 2	283	69			513	1443 •				
Dπ Ізму	194	157			139	318				
Ωπ ІюІеу	590	53			12	19				
Pt to ils t basin (Lt) handle	819	7056*/8007	11798 * / 159302 / 6067 / 5011	25	3274		156			
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Pttoiletfinch	٠		3504	i 17						
Pt Bathnoom shower tab	824		64 5	31	517		38			
Pt Bathnoom shower handle	3770 • / 2208		372	98	43		138			
bed 23A bedsile locler	376	219			1703					
bed 9A call bell	380			57						
bed 9A locker	348			30	122					
bed 10 siderail	382									
bed 10 BP suff	++1									
HO leyboard (Lt)	401									
HO leyboard (Rt)	289				319		125			
Remorts:										
• swab talen for est										
Post-cleansing means swabb	ed talen immediately	after cleaning with	outany contact by patient							
ATP passing result : less than 250										



- Cleaning work should be reinforced in targeted wards
- o Set basic standard
- Step up frequency of cleaning service for common touching surfaces
- o Observe compliance
- Demo cleaning method to those staff with lapse practice ASAP
- Collect feedback from users



瑪嘉烈醫院

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KOWLOON WEST CLUSTER

Evaluation and application of ATP detection as an effective audit tool for determining cleanliness of hospital environment surfaces and establishing appropriate benchmarks

Lam BHS¹, Tang WC², Ng TK^{1,2} ¹ Department of Pathology, Princess Margaret Hospital,²Infection Control Team, Princess Margaret Hospital

Background: These was specific dates thereing and time on the link between environmental contensates of vacuum contensate wave to appear. Every sense approximation of the specific dates and the specific da

Methods: 228 solution disk from 15 distance units (Academ 4 Emergency, Normaline Cam, Neptoniagy, Respiratory, Normaline), historikan Disson, Oncologi, Neurosciegiery, Orthosoeficia, Neuralita (Internationa Cam, Neuralita (Internativa) Cae, Neural



Selected wards and sites with unsatisfactory results, i.e. high or increased post-cleansing readings were repeated with ATP in only, and the cleansing staff of these words were notified to relefonce proper cleansing (Phase II).

Those failed again during Phase II were repeated with infection Control Nurses performing the cleansing procedure themselves and traded by ATC method again (Phase III)



Clean-Trace Study at PMH: Evaluation and application of ATP detection as an effective audit tool for determining cleanliness of hospital environment surfaces and establishing appropriate benchmarks

Lam BHS¹, Tang WC², Ng TK^{1,2}

¹Department of Pathology, Princess Margaret Hospital, ²Infection Control Team, Princess Margaret Hospital

Objectives

- To determine site-specific thresholds of ATP values as benchmarks for surface cleanliness by comparison of results of pre- and post-cleansing samples taken from common hand-touch surfaces in high-risk clinical areas.
- To show any correlation of the results by the ATP and ACC methods.
- To identify any highly contaminated environmental surface.
- To identify the extent of contamination by hospital pathogens such as MRSA and Acinetobacter spp.

Target areas

- Total 15 clinical areas
 - A&E
 - ONC-H4N
 - Renal-P2, P3
 - ICU-S6, F4
 - MID-S12
 - Resp-F3
 - Haem-C6
 - NSHDU-A6
 - O&T-D5
 - PICU-A1
 - PHaem-B1
 - NICU-D1
 - SHDU-D4

Phase I

- Sampling sites
 - sites under routine cleansing schedule: sampled twice for pre- and postcleansing testing on the same day (Total: 156 sites)
 - The cleansing procedures on the site sampled are supposed to be the standard ones or under supervision

Location 1: Bed unit

- Patient locker
- Bed table
- Bedside rail (or mattress for A&E)
- Suction port regulator
- BP meter cuff
- Touch screen of bedside monitor (for ICU, NSHDU, SHDU)
- Ventilator control panel (for Resp, ICU, NSHDU, SHDU)

Location 2: Nursing station

- Telephone handset
- CMS computer keyboard
- □ Location 3: Patient toilet
 - Toilet seat
 - Toilet flushing lever or button
 - Toilet door handle

- The sites for **spot check** will be sampled once only (Total: **70** sites)
 - □ Infusion pump
 - □ BP meter control panel
 - □ Stethoscope diaphragm
 - □ Tympanic thermometer
 - □ Trolley (dressing or multi-purpose)

Sampling methods

- **ATP**
 - Clean-Trace Surface swabs (3M, Brigend, UK)

 according to the manufacturer's instructions: swab the designated site of ~10cm x 10cm (or less for items of smaller size
 - Results (in relative light units, RLU) were read and recorded in the Clean-Trace NG luminometer
3M Clean-Trace - Sampling







- Clinical use dry swabs were moistened in sterile saline solution prior to use
- An area adjacent to and of the same size as the one swabbed by ATP method of the designated site
- swab suspended in 1ml sterile saline, vortexed for 10 s, 100µl on to a Columbia blood agar (BA) plate, CLED agar plate,1ml Mannitol Salt Broth (MSB), 1ml Acinetobacter Enrichment Broth (AEB) each
- The inoculated MSB and AEB were incubated for 24h at 30°C
- 10µl of MSB and AEB was then be subcultured to a MRSA chromogenic agar plate (BioMérieux, Marcy L'Etoile, France) and a Modified Leeds Acinetobacter Medium plate respectively
- All inoculated plates were incubated for 48 h at 35°C
- Any colony growth on the BA plates was counted (in colony forming units, CFU) and significant bacteria e.g. MRSA, *P. aeruginosa, Acinetobacter spp.* were looked for

Period for sampling

□ 3 – 23 September 2009

Phase II

- Selected wards and sites with unsatisfactory results, i.e. high or increased (as c.f. precleansing) post-cleansing readings, were repeated with ATP method only
- The cleansing staff of these wards were notified in advance to reinforce proper cleansing

Phase III

Those failed again during Phase II were repeated with ICT performing the cleansing procedure themselves and tested by ATC method again

Pre-/post-cleansing comparison



- Correlation between ATP & ACC
 Pre-cleansing: Pearson correlation = 0.254 (p = 0.000)
 Post-cleansing: Pearson correlation = 0.291 (p = 0.000)
 - □ Difference between pre-/post-cleansing: Pearson correlation = -0.10 (p = 0.907)

- Post-cleansing (all sites)
 ATP > 2.5 RLU/cm²: 86 (55%)
 - □ ACC > 10 CFU/cm²: 39 (25%)
- 22 (14%) sites required Phase II

ATP Post-readin	g (sites with d	ecreased readings only)		
Median		2.0000		
Std. Deviation		514.72309	No. of cases below the percentile	
Percentiles	50	2.0000	56 (out of 113)	
	60	5.1200	68 (out of 113)	
	70	7.9000	79 (out of 113)	
	75	11.6500	85 (out of 113) 91 (out of 113)	
	80	17.1200		
	85	21.9900	96 (out of 113)	
	90	34.6200	102 (out of 113)	
	95	138.3000	108 (out of 113)	
ACC Post-readin	ng (sites with d	ecreased counts only)		
Median Std. Deviation		.1000		
		13.47286	No. of cases below the percentile	
Percentiles	50	.1000	34 (ou of 93)	
	60	.2000	47 (out of 93)	
	70	.9000	64 (out of 93)	
	75	1.0000	66 (out of 93)	
	80	1.0200	75 (out of 93)	
	85	2.8200	79 (out of 93)	
	90	6.6000	84 (out of 93)	
	95	20 0000	88 (out of 93)	













Organisms:

□ 4 MRSA identified but the related patients were not MRSA carriers

Ward	Site	Pre-/Post- cleansing	ATP (RLU/cm²)	ACC (CFU/cm ²)
F4	Bed table	Pre	3.4	15
P2	BP meter cuff	Pre	13.2	15
F4	Telephone handset	Pre	76	11
C6	Toilet seat	Pre	1.9	37.5

Pseudomonas spp. and Acinetobacter spp. identified but not multidrug resistant

Spot check:
 ATP > 2.5 RLU/cm²: 60 (86%)
 ACC > 10 CFU/cm²: 16 (23%)

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1	Date 💌	Ward (N= 🖵	No. of sampling	bed no	Remarks T	pre-cleaning (AT) 🚽	post-cleaning (ATP 🔫	pre-cleaning (ATP) 🖵	post-cleaning (ATP)/cm2 🖵		
2			BP cuff(10cm×10cm)		SSD	32B	98	3.2B	0.9B		
3	23/11/2009	A&E	Patient locker ($10 \text{cm} \times 10 \text{cm}$)	Bed 9		B6	34	0.B6	0.34		
4			Toilet door handle (2cm × 5cm)			1054	14	105,4	1.4		
5			Patient locker (10cm × 10cm)	Bed 1P	ISS	755	Bl	7.55	0.81		
6		Al	Toilet door handle (2cm × 5cm)			62	49	62	49		
7		D1	Ventilator control panel(10cm × 10cm)	Bed 7N	ISS	435	18	4.35	0.1B		
8			Bedside rails (1cm ×10cm)	Bed 3	ISS	1366	57	13.66	0.57		
9		A6	Suction port regulator (1cm × 1cm)			1342	39	1342	39		
10			Bed table (10cm×10cm)		ISS	1576	217	15.76	2.17		
11	24/11/2009	C6	Toilet door handle (2cm × 5cm)			631	250	63.1	25		=
12			Patient locker (10cm × 10cm)		ISS	764	21	7.64	0.21		
13		F3	Bedside rails (1cm ×10cm)			1282	210	128.2	21		
14			toilet door handle (2cm × 5cm)			B49	21	B4.9	2.1		
15		F4	Suction port regulator (1cm × 1cm)		ICU staff	566	28	566	2B		
16		P2	Bedside rails (1cm ×10cm)		ISS	596	90	59.б	9		
17		15	Suction port regulator (1cm × 1cm)			5B3	21	5B3	21		
18	26/11/2009	S6	Suction port regulator (1cm × 1cm)		ICU staff	179	67	179	67		
19		610	Toilet flush lever(2cm × 2cm)		IDC staff	1147	17	286.75	425		
20		512	Toilet door handle (2cm × 5cm)			B73	13	87.3	1.3		
21			Bed table (10cm ×10cm)		ISS	250	19	19	0.19		
22		H4N	Toilet flush lever(2cm × 2cm)			2550	22	637.5	5.5		
23			Toilet door handle (2cm × 5cm)			1186	14	118.6	1.4		~
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■10 sites (45%) required Phase III

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	A	В	C No of sempling	D	E	F	G		^		
1	Date	Ward(N=9)	NO. OF Sampling	pre-cleaning (ATP)	post-cleaning (ATP)	pre-cleaning (ATP)/cm2	post-cleaning (ATP)/cm2				
2	30/12/2009	A1	Toilet door handle (2cm×5cm)	1945	10	194.5	1				
3		A6	Suction port regulator (1cm × 1cm)	100	3	100	3				
4		C6	Toilet door handle (2cm × 5cm)	100	9	10	0.9				
5		F3	Bedside rails (1cm×10cm)	278	16	27.8	1.6				
б		F4	Suction port regulator (1cm×1cm)	175	2	175	2				
7		P3	Bedside rails (1cm×10cm)	268	16	26.8	1.6				
8		P3	Suction port regulator (1cm × 1cm)	20	2	20	2				
9		S6	Suction port regulator (1cm × 1cm)	19	3	19	3				
10		S12	Toilet flush lever (2cm × 2cm)	640	5	160	1.25				
11		H4N	Bedside rails (1cm×10cm)	1676	15	167.6	1.5				
12		H4N	Toilet flush lever (2cm × 2cm)	1013	9	253.25	2.25				
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Discussion

- ATP method: rapid, objective, semi-quantitative, sensitive
- Linear correlation with ACC was not strong
 - □ different bacteria might release different amount of ATP
 - □ ATP can also be released from dust & organic matter but not viruses
 - □ Although not identifying specific microorganisms, its role could potentially be supplemented by ACC during outbreak investigation
- 2.5 RLU/cm² could generally be achievable and consistent with overseas standards except items with very small surface area (e.g. suction port regulator) for which 5 RLU/cm² should be reasonable
- Items with surface of rough fabrics were required to be cleaned more meticulously or microfibre cloth could be indicated though not tested in this study
- Items not under routine cleaning schedules showed highly unsatisfactory results
- Feedback to wards and cleansing staff of the results with reinforcement of proper cleansing showed improved results

Limitation

- Reproducibility was not tested (operator dependent)
- Tracking of MRSA isolated was not performed

Conclusion

- ATP method as a real-time, user-friendly, sensitive assessment or audit tool
- Comparable benchmarks could be established as with overseas standards
- Items with rough fabrics require special attention for deep cleaning
- Good cleaning practice cannot be overemphasized
- Regular cleaning schedule should be extended to other items with potential spread of microorganisms to patients and healthcare workers

What you see is NOT what you get!

Ideal environmental disinfectant

Efficacious

- Broad spectrum
- Great log reduction
- Long lasting with residual effect
- Not operator-dependent
- Not inducing microbial resistance
- Not affected by organic matter
- Efficient, reliable, penetrative
- Stable, durable
- > User friendly, non-odorous
- Non-interruptive to cleansing routine
- Compatible with various surface materials
- Safe
- Environmentally friendly
- Short TAT
- Inexpensive



K.L. Yeung



Joseph Kwan



Arthur Lau



Smart Anti-Microbial Coating

King Lun Yeung¹, Joseph Kwan^{2,3}, Arthur P.S. Lau²

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¹Tel.: 2358 7123, Fax: 2358 0054, E-mail: kekyeung@ust.hk

Smart Anti-Microbial Coatings

Anatomy of the Multilevel Anti-Microbial Formulation





Courtesy of Prof. King L. Yeung, HKUST

Fast and Effective

Sprayed-on Formulation

≥ 99.99 % within 1 min

Coating on Surface

Bacteria: 99.9 % within 1 min H1N1 human swine flu virus: 99 % within 3 min Bacillus spores: 99 % within 30 min



Long Lasting

Minimum 30 days in single application



Effectiveness can be maintained for at least <u>60</u> days after coating



Courtesy of Prof. King L. Yeung, HKUST

SMART

Courtesy of Prof. King L. Yeung, HKUST

Persistent Release of Biocides





Disinfect up to 3 mm from coating

Time (days)





SMART

Courtesy of Prof. King L. Yeung, HKUST

Rapid Self-disinfection



Contact Time (min)

Safe and Environmentally Friendly

All active formulations are U.S. FDA and EPA approved;

Smart dosing ensures long-term safety and prevents resistance in microorganisms;

Environmentally friendly;

Biodegradable





No skin reaction

Courtesy of Prof. King L. Yeung, HKUST

Safe and Compatible

Courtesy of Prof. King L. Yeung, HKUST



No detectable CIO2 (i.e., < 0.01 ppm)

Safe and Compatible

Tested <u>compatible</u> with stainless steel, aluminum, plastics, paper, cotton...

Tested durable under repeated wiping with

- Dry cotton cloth and napkin paper
- Wet cotton cloth with tap water
- Wet cotton cloth with 1:49 bleach solution
- Wet cotton cloth with Virkon disinfectant solution





Collaborative studies with HKUST

- Provision of MRSA, multi-drug resistant *P. aeruginosa, Acinetobacter spp.,* enterococci strains for testing at HKUST lab (2010)
- 2. Testing with MRSA and Acinetobacter spp. (Efficacy Test, Persistent Test, Soiling Test, Coating Durability under Standard Cleaning) on bed table at PMH lab (Mar 2011)

PMH Field Test

May 11 to 26, 2011

Dr. T K Ng Dr. Bosco Lam

Princess Margaret Hospital



Objectives:

Coating efficacy and persistence against MDRO.

Lab tests against MDRO



Courtesy of Prof. King L. Yeung, HKUST *

Laboratory Test

Courtesy of Prof. King L. Yeung, HKUST



0.1ml of 10⁶/ml of MRSA on glass with contact time of 10 min





Field Test

May 11 to 26, 2011

Objectives:

Coating efficacy and persistence against MDRO.



Coating applied by wiping, let dry 1 day Challenged by drop test of 10⁵ organisms / grid Sample 5 grids per day



140 2"x2" grids coated with 0.1 g coating each

Courtesy of Prof. King L. Yeung, HKUST

Coating Efficacy

Long-term Study of Coating Performance (12 days)



Challenged with 10⁵ MDRO and sampling by swabbing after 10 min contact. Please note reduction is calculated against control surface without coating.

Courtesy of Prof. King L. Yeung, HKUST
Coating Efficacy

Repeated Challenge and Surface Soiling



Challenged with 10⁵ MDRO twice daily and sampling by swabbing after 30 min contact. Please note reduction is calculated against control surface without coating.

Courtesy of Prof. King L. Yeung, HKUST

Collaborative studies with HKUST

- 3. Field tests at clinical surfaces (high-touch) and equipments at IDC simulation ward
- Clinical trials on contamination / infection rates in general wards / LKB / ICU

Room Decontamination by Hydrogen Peroxide Vapor (HPV)



- > 10-30%: bactericidal, virucidal, sporicial
- Disinfectant for inanimate materials & inert surfaces free of catalase
- Solutions: for surgical implant components, temp.-sensitive plastic equipment, spacecraft hardware, hydrophilic soft contact lenses, commercial packing materials, water, milk
- HPV: for pharmaceuticals, foodstuffs, processing equipment, packaging materials, fermentors, dialyzers, incubators, lyophilizers, BSC, glove boxes, centrifuges

Types of HPV

Micro-condensation

- Bioquell, UK
- Gas (fumigation), 30% H_2O_2 , <1 μ m
- High humidity
- Broken down catalytically to water vapour & oxygen
- Require sealing of all ventilation ducts, windows, doors
- Need checking for leakage (safety monitoring)
- Need biological indicators (spore strips)
- Dry mist
 - Sterinis (Gloster, France→Glosair, J&J, UK); VaproSure (Steris), USA
 - Aerosol (fogging), 5% H₂O₂, <50ppm phosphoric acid, <50ppm silver cations, 8-12μm
 - Low humidity
 - Decompose spontaneously
 - Room sealing not necessary
 - checking for leakage not needed (?)
 - No BI
 - Boyce. Infect Control Hosp Epidemiol 2009;30;515-7
 - Otter et al. Infect Control Hosp Epidemiol 2010;31:1201-2
 - Pottage et al. J Hosp Infect 2010;74:55-61

Field tests at PMH (30 Mar – 1 Apr 2009)

Challenge with Acinetobacter spp. and MRSA at 2.0 McFarland standard (~6x10⁸CFU/ml) before HPV fumigation

Comparing before and after by

- ATP
- Culture
- Location:
 - D2 (unused ICU)
 - S8 (isolation ward)
 - BSL3 Lab
 - Cold chamber at Mortuary

Step 1. Seal up the room exhaust to building corridor



S8 Ward



S8 Ward

S8 Ward



Step 2. Place the H2O2 Generator at the center of the Isolation Room. Open the Pass Through Hatch door and place the fan blowing to the bathroom.



Step 3. Open the ceiling plates, prepare to disinfect the HEPA Filter



Step 4. Place the Control Panel at outside, set up program, insert H2O2 volume parameter and start cycle. The H2O2 Vapour will fill up in the Isolation Room

The total Cycle time around 3-4 hours

Princess Margaret Hospital – General Ward D5



Step 1. Seal up the air-conditional exhaust and the gap between wall and false ceiling









Step 2. Place the H2O2 Generator and R20 Aeration Module at the ocnter of the General Ward



Step 3. Place the Control Panel at outside, set up program, insert H2O2 volume and start cycle. Total Cycle time around 3-4 hours.

Biological Indicator (spore) – for gas sterilization



ATP Test & Swab culture pre and post fumigation at D2 (bed 10-13)

			ACIS 2.0			MRSA 2.0			
No	Location	Organicm		Count	Count		Count	Count	
NO	Location	organishi	ATP	on BA	on MAC	ATP	on BA	on MAC	
				cfu	cfu		cfu	cfu	
1	Suction port regulator	Pre	939	30	40	727	NG	NG	
		Post	4	NG	NG	3	NG	NG	
2	Computer stand	Pre	285	30	36	1726	1	0	
		Post	47	NG	NG	98	NG	NG	
3	Monitor	Pre	672	26	62	1465	0	1	
		Post	153	NG	NG	199	NG	NG	
4	Wall bench	Pre	380	100-200	90	1472	NG	NG	
		Post	11	1	NG	33	NG	NG	
5	Window	Pre	172	NG	NG	1002	2	1	
		Post	4	NG	NG	6	NG	NG	
6	Fan	Pre	289	1	6	2059	1	2	
		Post	33	NG	NG	637	NG	NG	
7	Keyboard box	Pre	468	NG	NG	684	0	3	
		Post	162	NG	NG	266	NG	NG	
8	Keyboard	Pre	1098	100-200	100-200	879	4	4	
		Post	132	NG	NG	92	NG	NG	
9	Lamp switch	Pre	1173	2	5	1419	9	5	
		Post	121	NG	NG	158	NG	NG	
10	Tap under sink	Pre	567	2	5	618	15	14	
		Post	81	NG	NG	320	NG	NG	



ATP & Swab culture pre and post fumil	ligation at S8 isolation room 1/4/200
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			ACIS 2.0			MRSA 2.0			
No	Location	Organicm		Count	Count		Count	Count	
	Location	organism	ATP	on BA	on MAC	ATP	on BA	on MAC	
				cfu	cfu		cfu	cfu	
1	Electricity socket	Pre	1701	60	~ 100	79	13	3	
		Post	27	NG	NG	19	NG	NG	
2	Suction port regulator	Pre	940	60	60	207	10	7	
		Post	100	NG	NG	60	NG	NG	
3	Call bell button	Pre	1617	50	50	148	15	8	
		Post	53	NG	NG	20	1	NG	
4	Low level air exhaust	Pre	1271	50	70	293	2	3	
		Post	86	NG	NG	27	NG	NG	
5	Toilet sink	Pre	3482	80	100-200	411	15	24	
		Post	25	NG	NG	26	NG	NG	
6	Toilet flush button	Pre	2648	60	60	453	70	60	
		Post	118	NG	NG	13	NG	NG	
7	Hatch	Pre	162	70	80	584	60	40	
		Post	26	NG	NG	13	NG	NG	
8	Toilet door handle	Pre	851	~ 100	~ 100	274	17	10	
		Post	246	NG	NG	20	NG	NG	
9	Curtain rail	Pre	570	27	23	352	40	10	
		Post	20	NG	NG	22	NG	NG	
10	Auto door sensor	Pre	4465	80	80	343	40	40	
		Post	71	NG	NG	23	NG	NG	

ATP Test pre and post fumigation

Date: 30/03/2009

Location: S16 BSL 3

No	Location	MRS	A 2.0	ACIS 2.0		
NO	Location	Pre	Post	Pre	Post	
1	Incubator	875	22	471	17	
2	Telephone	806	32	282	73	
3	Cupboard under sank	600	105	291	171	
4	Water tape on floor	710	18	111	18	
5	Autoclaver	612	29	500	40	
6	BSC	2128	22	90	19	

Agar plate result after fumigation:

No	Agar plate location	Organisms	ATP count
1	Sank	PAER	39941
2	Incubator	ACIS	21013
3	BSC	MRSA	76994

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ATP & Swab culture pre and post fumigation at Mortuary

Date: 1/4/2009

Location: Mortuary

		Organism		ACIS 2.0		MRSA 2.0		
No	Location		ATP	Count on BA cfu	Count on MAC cfu	ATP	Count on BA cfu	Count on MAC cfu
1		Pre	3172	60	60	225	11	4
		Post	15	NG	NG	14	NG	NG
2	Inside the cold chamber	Pre	1073	80	45	1068	29	22
		Post	126	NG	NG	14	NG	NG
3		Pre	1700	60	~ 100	642	28	23
		Post	22	NG	NG	14	NG	NG
4		Pre	2779	50	~ 100	145	10	20
		Post	45	NG	NG	23	NG	NG
5		Pre	656	100 - 200	~ 100	161	45	35
		Post	17	NG	NG	16	NG	NG

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Applications at PMH

- Interim disinfection for cubicles or rooms with uncontrolled spread of MDRO (e.g. MRSA, MRAB) and problematic pathogens (e.g. C. difficile, Norovirus, VRE).
- Terminal disinfection for cubicles or rooms with highly pathogenic or bioterror agents (e.g. smallpox, plague).
- Post-renovation (with much dust generation) disinfection for wards housing immunocompromised or highly susceptible inpatients or with high prevalence of MDRO.

- what's the magnitude of the problem of your setting (incidence? Surveillance in place?)
- Environmentally hardy pathogens (higher priority than CRE)
- Not for routine terminal disinfection for endemic pathogens
 - may be considered for single room previously occupied by VRE, C. difficile ribotype 027 patient (?)
- Source control / removal
- Have all other IC measures / cleansing standard been optimized and adhered?
- Factors modifiable?
- Take any chance to vacate the room / cubicle / ward?

Any ways to avoid re-contamination of the fumigated ward

- MRSA:
 - Decolonize known MRSA carriers before moving them back to the ward or relocate them to other cohorted areas
 - Perform active screening and prompt isolation of newly identified MRSA carriers
- Take chance of decontamination of selected medical equipment (non-critical items; semi-critical items??), not limiting to those belong to the ward for fumigation
 - French et al. J Hosp Infect 2004;57:31-7
 - But decontamination of narrow-lumen items should be done by appropriate standard methods
 - Otter et al. J Hosp Infect 2006;62:384-92

- Give ample time (at least 2-4 weeks) for all relevant parties (EMSD, FM, ICT, dept & ward manager, lab technologist) for joint site inspection and planning if any special preparation / remedial work (renovation) is required & feasibility of setting up the venue
- Total cost largely depends on size of the area / no. of sessions applicable for HPV
- Pre-cleansing is mandatory

> How to measure effectiveness?

- Environmental sampling, before and after
 - Specimens? Frequent hand-touch sites only?
 - Culture? ATP? But not fluorescent markings.
 - Typing (PFGE)
- Clinical outcomes
 - e.g. MRSA infection / colonization rate; acquisition rate (admission screening and FU screens)
- Not to replace routine standard environmental decontamination

Other applications

> lab

- BSC
- Autopsy room / mortuary: cold chamber for dead bodies
- Centrifuge (need injection port)
- Isolation ward
 - HEPA unit



To buy or not to buy...



More innovative products...





Thank you!